**INSTRUCTIONS**

**The Principal Investigator is responsible for correctly classifying their activities.**

**Use the checklist below to determine whether registration is required:**

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| **Does your activity involve:** | |  |
| i | Molecules that   1. Are constructed by joining nucleic acid molecules, and 2. That can replicate in a living cell, i.e., recombinant nucleic acids? | No  Yes |
| ii | Nucleic acid molecules that are chemically or by other means synthesized or amplified, including those that are chemically or otherwise modified but can base pair with naturally occurring nucleic acid molecules, i.e., synthetic nucleic acids? | No  Yes |
| iii | Molecules that result from the replication of those described in (i) or (ii) above? | No  Yes |

**If you answered NO to ALL of the above = STOP! Your activity does not require IBC review. Contact Environmental Health & Safety at** [**EHS@baylor.edu**](mailto:EHS@baylor.edu) **prior to beginning the activity in your laboratory.**

**If you answered YES, to ANY of the above = continue with this form.**

**For questions about this form or the IBC, contact** [**IBC@baylor.edu**](mailto:IBC@baylor.edu)

**Submit this form to** [**IBC@baylor.edu**](mailto:IBC@baylor.edu)**.**

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| **PROJECT TITLE** | Click here to enter text. |

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| **SUBMISSION TYPE** | New Application  3-year Renewal IBC# Click here to enter text. |
| **CLASSIFICATION** | Research  Teaching |

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| **PRINCIPAL INVESTIGATOR** | | |
| Name: Click here to enter text. | | |
| Department: Click here to enter text. | | |
| Email: Click here to enter text. | | Phone: Click here to enter text. |
| Faculty  Staff  Student | | |
| **IF STUDENT:** | | |
| Graduate/Professional Student  Degree Program: Click here to enter text. | Undergraduate Student  Degree Program: Click here to enter text. | |
| Faculty Advisor: Click here to enter text. | | |
| Email: Click here to enter text. | Phone: Click here to enter text. | |

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| **LABORATORY CONTACT (other than PI)** | |
| Name: Click here to enter text. | |
| Title: Click here to enter text. | |
| Email: Click here to enter text. | Phone: Click here to enter text. |

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| **LOCATIONS & PERSONNEL** | | | | | | | |
| 1. | | **List all Baylor locations where work will be conducted in the table below:** | | | | | |
| Lab | Building(s) | | Room Number(s) | | |
| Click here to enter text. | Click here to enter text. | | Click here to enter text. | | |
| Click here to enter text. | Click here to enter text. | | Click here to enter text. | | |
| Click here to enter text. | Click here to enter text. | | Click here to enter text. | | |
| Click here to enter text. | Click here to enter text. | | Click here to enter text. | | |
| 2. | | Will you be conducting work at or collaborating with any institutions external to Baylor but within the United States?  If YES, identify the institution and the work to be conducted: Click here to enter text. | | | | No  Yes | |
| 3. | | Will you be conducting work at or collaborating with any institutions outside the United States?  If YES, identify the institution and the work to be conducted: Click here to enter text. | | | | No  Yes | |
| 4. | | If you are collaborating with a researcher outside of Baylor, has the researcher been reviewed by their institution’s IBC?  If YES, Baylor IBC may request a copy of their IBC’s letter of approval. | | | | No  Yes  N/A | |
| 5. | **List all individuals working on this protocol other than the PI.** Include internal collaborators, technicians, post docs, graduate students, work study students, volunteers, etc. *Attach a separate page if necessary.* | | | | | |
| **Name** | | **Study Responsibilities** | | | |
| Click here to enter text. | | Click here to enter text. | | | |
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| Click here to enter text. | | Click here to enter text. | | | |
| 6. | Does your proposed activity involve information, technology, materials or intellectual property that has been deemed to be sensitive or protected against open publication or disclosure, i.e. classified, proprietary, business sensitive, sponsor restricted, etc.  If YES, you may be subject to Federal Export Controls Regulations. | | | | | No  Yes |
| 7. | Does the project involve transferring organisms or genetic material between Baylor and another entity?  If yes, a Material Transfer Agreement may be required. Contact Research Compliance. | | | | | No  Yes |
| 8. | Will you be importing or exporting organisms or genetic material which requires a permit or license?  If yes, export controls may apply. Contact Research Compliance. | | | | | No  Yes |

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| **PROJECT DESCRIPTION** | | | | |
| 9. | In lay language, provide a summary of your overall objectives of this project:  Click here to enter text. | | | |
| 10. | Summarize the purpose, goals, and anticipated outcomes:  Click here to enter text. | | | |
| 11. | If you are submitting a 3-year renewal, provide a summary report of the last 3 years working on this protocol:  N/A  Click here to enter text. | | | |
| 12. | Provide a step-by-step ‘walk-through’ of your research methodology and the rationale behind the choice of experiments toward your goal:  Click here to enter text. | | | |
| 13. | Explain why and how specific r/sNA materials are used:  Click here to enter text. | | | |
| 14. | State previous work experience with the r/sNA and/or procedures specified in this project.  Click here to enter text. | | | |
| 15. | Funding Source(s)  External Funding/Subcontract – Identify below  No Funding  Internal Funding | | | |
| Funding Agency | Are you the primary awardee: | Have you subcontracted any part of this grant? If yes, identify the institution | |
| Click here to enter text. | No  Yes | No  Yes: Click here to enter text. | |
| Click here to enter text. | No  Yes | No  Yes: Click here to enter text. | |
| Click here to enter text. | No  Yes | No  Yes: Click here to enter text. | |
| Click here to enter text. | No  Yes | No  Yes: Click here to enter text. | |
| 16. | Will this research utilize viable organisms containing r/sNA in culture volumes of 10 liters or greater? | | | No  Yes |

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| **NIH GUIDELINES** | | | |
| Indicate the experiment or manipulation of materials subject to the NIH Guidelines for Research Involving Recombinant or Synthetic Nucleic Acid Molecules (*NIH Guidelines*). For recombinant and synthetic nucleic acids, indicate the relevant Section of the Guidelines and the appropriate Biosafety Level (BSL) according to the [*NIH Guidelines*](https://osp.od.nih.gov/biotechnology/nih-guidelines/), [CDC/NIH Biosafety in Microbiological and Biomedical Laboratories](https://www.cdc.gov/labs/BMBL.html) (BMBL), and/or University policies. The University Biosafety Officer can assist researchers with this.  **Reminder: The Principal Investigator is responsible for the correct classification of their activities!** | | | |
| 17. |  | [Section III-A-1-a](https://osp.od.nih.gov/wp-content/uploads/2019_NIH_Guidelines.htm#_Toc3457033) | Deliberate transfer of a drug resistance trait to microorganisms that are not known to acquire the trait naturally. (Do not check for standard drug resistance, e.g., ampicillin into E. coli.) |
| 18. |  | [Section III-B-1](https://osp.od.nih.gov/wp-content/uploads/2019_NIH_Guidelines.htm#_Toc3457035) | Experiments involving the cloning of toxin molecules with LD50 of less than 100 nanograms per kilogram body weight. |
| 19. |  | [Section III-B-2](https://osp.od.nih.gov/wp-content/uploads/2019_NIH_Guidelines.htm#_Toc3457036) | Experiments that have been approved as Major Actions under the *NIH Guidelines*. |
| 20. |  | [Section III-D-1](https://osp.od.nih.gov/wp-content/uploads/2019_NIH_Guidelines.htm#_Toc3457040) | Experiments using Risk Group 2, 3, or 4, or restricted agents as host-vector systems. If applies, check subcategory:  a  b  c  d |
| 21. |  | [Section III-D-2](https://osp.od.nih.gov/wp-content/uploads/2019_NIH_Guidelines.htm#_Toc3457041) | Experiments in which DNA from Risk Group 2, 3, or 4 agents or restricted organisms is cloned into nonpathogenic prokaryotic or lower eukaryotic host vector systems. |
| 22. |  | [Section III-D-3](https://osp.od.nih.gov/wp-content/uploads/2019_NIH_Guidelines.htm#_Toc3457042) | Experiments involving the use of infectious DNA or RNA viruses, or defective DNA or RNA viruses in the presence of helper virus in tissue culture systems. If applies, check subcategory:  a  b  c  d  e |
| 23. |  | [Section III-D-4](https://osp.od.nih.gov/wp-content/uploads/2019_NIH_Guidelines.htm#_Toc3457043) | Experiments involving whole animals in which the animal’s genome has been altered by stable introduction of recombinant or synthetic nucleic acid molecules, or nucleic acids derived therefrom, into the germ-line (transgenic animals) and experiments involving viable recombinant or synthetic nucleic acid molecule-modified microorganisms tested on whole animals. If applies, check subcategory:  a  b  c-1  c-2 |
| 24. |  | [Section III-D-5](https://osp.od.nih.gov/wp-content/uploads/2019_NIH_Guidelines.htm#_Toc3457044) or [Section III-E-2](https://osp.od.nih.gov/wp-content/uploads/2019_NIH_Guidelines.htm#_Toc3457049) | Experiments involving whole plants. If applies, check subcategory:  Section III-D-5-  a  b  c  d  e / Section III-E-2-  a  b-1  b-2  b-3  b-4  b-5 |
| 25. |  | [Section III-D-6](https://osp.od.nih.gov/wp-content/uploads/2019_NIH_Guidelines.htm#_Toc3457045) | Experiments involving more than 10 liters of culture. |
| 26. |  | [Section III-D-7](https://osp.od.nih.gov/wp-content/uploads/2019_NIH_Guidelines.htm#_Toc3457046) | Experiments involving Influenza Viruses. If applies, check subcategory:  a  b  c  d |
| 27. |  | [Section III-E-1](https://osp.od.nih.gov/wp-content/uploads/2019_NIH_Guidelines.htm#_Toc3457048) | Experiments involving the formation of rDNA molecules containing no more than two-thirds of the genome of any eukaryotic virus (except RG3, 4 or restricted agents) when performed in tissue culture in the absence of helper virus. |
| 28. |  | [Section III-E-2](https://osp.od.nih.gov/wp-content/uploads/2019_NIH_Guidelines.htm#_Toc3457049) | See number 24 above. |
| 29. |  | [Section III-E-3](https://osp.od.nih.gov/wp-content/uploads/2019_NIH_Guidelines.htm#_Toc3457050) | Experiments involving transgenic rodents. This section covers experiments involving the generation of rodents in which the animal's genome has been altered by stable introduction of recombinant or synthetic nucleic acid molecules, or nucleic acids derived therefrom, into the germ-line (transgenic rodents).  Check here is Section III-E-3-a applies: Experiments involving the breeding of certain BL1 transgenic rodents are exempt under Section III-F, Exempt Experiments (See Appendix C-VIII, Generation of BL1 Transgenic Rodents via Breeding). |
| 30. |  | [Section III-F-1](https://osp.od.nih.gov/wp-content/uploads/2019_NIH_Guidelines.htm#_Toc3457051) | Those synthetic nucleic acids that: (1) can neither replicate nor generate nucleic acids that can replicate in any living cell (e.g., oligonucleotides or other synthetic nucleic acids that do not contain an origin of replication or contain elements known to interact with either DNA or RNA polymerase), and (2) are not designed to integrate into DNA, and (3) do not produce a toxin that is lethal for vertebrates at an LD50 of less than 100 nanograms per kilogram body weight. If a synthetic nucleic acid is deliberately transferred into one or more human research participants and meets the criteria of Section III-C, it is not exempt under this Section. |
| 31. |  | [Section III-F-2](https://osp.od.nih.gov/wp-content/uploads/2019_NIH_Guidelines.htm#_Toc3457051) | Those that are not in organisms, cells, or viruses and that have not been modified or manipulated (e.g., encapsulated into synthetic or natural vehicles) to render them capable of penetrating cellular membranes. |
| 32. |  | [Section III-F-3](https://osp.od.nih.gov/wp-content/uploads/2019_NIH_Guidelines.htm#_Toc3457051) | Those that consist solely of the exact recombinant or synthetic nucleic acid sequence from a single source that exists contemporaneously in nature. |
| 33. |  | [Section III-F-4](https://osp.od.nih.gov/wp-content/uploads/2019_NIH_Guidelines.htm#_Toc3457051) | Those that consist entirely of nucleic acids from a prokaryotic host, including its indigenous plasmids or viruses when propagated only in that host (or a closely related strain of the same species), or when transferred to another host by well-established physiological means. |
| 34. |  | [Section III-F-5](https://osp.od.nih.gov/wp-content/uploads/2019_NIH_Guidelines.htm#_Toc3457051) | Those that consist entirely of nucleic acids from a eukaryotic host including its chloroplasts, mitochondria, or plasmids (but excluding viruses) when propagated only in that host (or a closely related strain of the same species). |
| 35. |  | [Section III-F-6](https://osp.od.nih.gov/wp-content/uploads/2019_NIH_Guidelines.htm#_Toc3457051) | Those that consist entirely of DNA segments from different species that exchange DNA by known physiological processes, though one or more of the segments may be a synthetic equivalent. A list of such exchangers will be prepared and periodically revised by the NIH Director after appropriate notice and opportunity for public comment (see Section IV-C-1-b-(1)-(c), Major Actions). See Appendices A-I through A-VI, Exemptions under Section III-F-6--Sublists of Natural Exchangers, for a list of natural exchangers that are exempt from the NIH Guidelines. |
| 36. |  | [Section III-F-7](https://osp.od.nih.gov/wp-content/uploads/2019_NIH_Guidelines.htm#_Toc3457051) | Those genomic DNA molecules that have acquired a transposable element, provided the transposable element does not contain any recombinant and/or synthetic DNA. |
| 37. |  | [Section III-F-8](https://osp.od.nih.gov/wp-content/uploads/2019_NIH_Guidelines.htm#_Toc3457051) | Those that do not present a significant risk to health or the environment (see Section IV-C-1-b-(1)-(c), Major Actions), as determined by the NIH Director following appropriate notice and opportunity for public comment. See Appendix C, Exemptions under Section III-F-8 for other classes of experiments which are exempt from the NIH Guidelines. |
| 38. |  | Click here to enter text. | Add the section(s) or appendices of NIH Guidelines that apply to your experiments if not listed. If applies, add the section/appendix number(s) in the column to the left and provide explanation: Click here to enter text. |

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| **NATURE OF INSERTED OR CLONED MATERIAL (  N/A)** | | | | | | |
| 39. | Provide information below on the nature and species of sequence(s) added. | | | | | |
| Name of Sequence | Source  (species name) | Type of insert (knock-in, knock-out, point mutation, siRNA, transgene, etc.) | Function/impact on cell (ex: cell cycle regulator, oncogenic) | Promoter | |
| Click here to enter text. | Click here to enter text. | Click here to enter text. | Click here to enter text. | Click here to enter text. | |
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| Click here to enter text. | Click here to enter text. | Click here to enter text. | Click here to enter text. | Click here to enter text. | |
| 40. | Identify the cloning/expression/transfection vector(s) that will be used and provide a restriction map of vector unless it is commercially available. If commercially available, indicate vendor and catalog number. Include hyperlink, if possible.  Click here to enter text. | | | | | |
| 41. | Indicate method of gene transfer. Examples include transfection (eukaryotes), transformation (prokaryotes), electroporation, viral transduction. For viral transduction, fill out Section “Viral Vectors” below.  Click here to enter text. | | | | | |
| 42. | Do any sequences encode for a toxin? | | | | | No  Yes |
| If yes, provide information on these specific sequences. If a toxin has been modified to render it less toxic or nontoxic, explain and provide references that demonstrate this.  Click here to enter text. | | | | | |
| 43. | Identify the recipient host: Click here to enter text. | | | | | |

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| **VIRAL VECTORS (  N/A)** | | | | | | |
| 44. | List all viral vectors to be used and indicate any safety features that prevent generation of a replication competent product. | | | | | |
| Type of viral vector (e.g., lentivirus, adenovirus, AAV) | Expression plasmid  (i.e., transfer vector) | Packaging System | Safety Features  (e.g., self-inactivating) | Promoter driving transgene expression | |
| Click here to enter text. | Click here to enter text. | Click here to enter text. | Click here to enter text. | Click here to enter text. | |
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| Click here to enter text. | Click here to enter text. | Click here to enter text. | Click here to enter text. | Click here to enter text. | |
| 45. | If purchasing pre-made viral particles, indicate vendor and catalog number. Include hyperlinks to products, if possible.  Click here to enter text. | | | | | |
| 46. | Is testing for replication competent virus required? (Testing is usually not required for self-inactivating viral vectors.) | | | | | No  Yes |
| If yes, indicate testing method or provide documentation if the vendor performs testing. A log of the results must be made available upon request. If replication competence is found, the Biosafety Officer must be notified.  Click here to enter text. | | | | | |

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| **GENE EDITING TECHNOLOGY (  N/A)** | | | | | |
| Complete this section if you are using gene editing technology (CRISPR, TALENs, zinc fingers, etc.). | | | | | |
| 47. | Describe the editing system (CRISPR, TALENs, zinc fingers, etc.).  Click here to enter text. | | | | |
| 48. | Indicate the organism to be edited. Indicate whether the editing is restricted to cell cultures or if it will result in edited whole animals or plants.  Click here to enter text. | | | | |
| 49. | Will genes coding for a functional CRISPR system (or similar gene editing tool) be stably integrated into the genome of the organism to create a transgenic strain? | | | | No  Yes |
| 50. | Does this CRISPR system have one or more RNA sequences (guide RNAs) that can target the endonuclease to cut regions of the homologous wild-type locus corresponding to insertion of any of the CRISPR components? | | | | No  Yes |
| 51. | Can this transgenic organism reproduce sexually? | | | | No  Yes |
| 52. | If editing whole animals or plants, will germline cells be targeted?  If yes, please address the potential to create a gene drive. Include safeguards to contain the gene drive, or safeguards to prevent creation of a gene drive.  Click here to enter text. | | | | No  Yes  N/A |
| 53. | How are the components (e.g. Cas9 and sgRNA) delivered? Common examples are plasmid transfection, viral vector, or nanoparticle delivery of purified components.  Click here to enter text. | | | | |
| 54. | Are the components (e.g. Cas9 and sgRNA) encoded by the same DNA construct or by multiple constructs? Provide the construct information, such as plasmid maps.  Click here to enter text. | | | | |
| 55. | Target gene function: Provide information on the genes that will be edited. | | | | |
| Gene name | Original gene function | Nature of modification | Anticipated impact on gene function | |
| Click here to enter text. | Click here to enter text. | Click here to enter text. | Click here to enter text. | |
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| 56. | Is the targeting mechanism specific to animals, humans, or could it affect both?  Click here to enter text. | | | | |
| 57. | What is known about off-target effects with the system components you are using?  Click here to enter text. | | | | |
| 58. | Indicate steps to minimize off-target effects. Examples include but are not limited to use of specific CRISPR design tools, or modified endonucleases with enhanced specificity.  Click here to enter text. | | | | |

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| **USE, ACQUISITION, OR CREATION OF TRANSGENIC ORGANISMS** | | | | | | | |
| * Completion of this section is required for the use, acquisition (transfer, purchase, etc.), or creation of transgenic invertebrates, vertebrates, or plants. * Exempt from this section is the acquisition, use, or breeding or transgenic **rodents** as long as they **do not** contain the following genetic modifications:  1. Incorporation of more than one-half of the genome of an exogenous eukaryotic virus from a single family of viruses; or 2. Incorporation of a transgene that is under the control of a gammaretroviral terminal repeat (LTR); or 3. Any transgenic rodent that results from this breeding is not expected to contain more than one-half of the genome of an exogenous eukaryotic virus from a single family of viruses. | | | | | | | |
| 59. | Will you be creating transgenic organisms, breeding transgenic organisms, exposing organisms to r/sNA, or purchasing/obtaining transgenic organisms from a commercial vendor or collaborator?  If NO or EXEMPT, skip the rest of this section.  If YES, explain: Click here to enter text. | | | | Yes  No  Rodent breeding exempt | | |
| 60. | Describe the transgenic organism(s) and how they will be used. If creating a new organism, describe the methods to be used.  Click here to enter text. | | | | | | |
| 61. | Genetic Modification Information | | | | | | |
| Name of inserted or modified genetic material | | Impact of genetic modification | Method used to introduce modification | Recipient strain or organism | | |
| Click here to enter text. | | Click here to enter text. | Click here to enter text. | Click here to enter text. | | |
| Click here to enter text. | | Click here to enter text. | Click here to enter text. | Click here to enter text. | | |
| Click here to enter text. | | Click here to enter text. | Click here to enter text. | Click here to enter text. | | |
| Click here to enter text. | | Click here to enter text. | Click here to enter text. | Click here to enter text. | | |
| 62. | Does the inserted genetic material encode a toxin or other hazardous agent?  If yes, describe: Click here to enter text. | | | | | | Yes  No |
| 63. | Genetically modified organisms cannot be released into the environment. Describe methods to prevent release (e.g., fly trap, specialized housing, etc.) and methods for disposal of these organisms:  Click here to enter text. | | | | | | |
| 64. | IACUC Status ( N/A – IACUC not required) | | | | | | |
| Protocol # | Click here to enter text. | | | | | |
| Status | Approved  Pending Approval  Not submitted | | | | | |
| 65. | Are any USDA APHIS permits or other regulatory permits required? | | | | | No  Yes | |

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| **RISK GROUP, BIOSAFETY LEVEL, SPECIAL CONTAINMENT/FACILITIES** | | | |
| Use the links to the resources listed below to:   1. Classify the risk group (Risk Group 1-3\*) for the work being conducted on this protocol AND 2. Determine the level of containment and procedures for this protocol (BSL-1-3\*).   NOTE: Risk group (RG) must be assigned when applicable. Not all experiments may be listed in the guidelines or on the websites listed below. Assign the highest level if declaring multiple agents. Biosafety level (BSL) must be assigned. **\* Work with RG-3 agents or in the BSL-3 laboratory requires prior consultation with the IBC, EHS, BSO, and Research Compliance.**  BMBL: <https://www.cdc.gov/labs/pdf/CDC-BiosafetyMicrobiologicalBiomedicalLaboratories-2009-P.PDF>  CDC Select Agents List: [www.selectagents.gov/SelectAgentsandToxinsList.html](http://www.selectagents.gov/SelectAgentsandToxinsList.html)  NIH Guidelines: <https://osp.od.nih.gov/wp-content/uploads/2019_NIH_Guidelines.htm>  ABSA Risk Group Database: <https://my.absa.org/tiki-index.php?page=Riskgroups>  Pathogen Safety Data Sheets and Risk Assessment: [www.phac-aspc.gc.ca/lab-bio/res/psds-ftss/index-eng.php](http://www.phac-aspc.gc.ca/lab-bio/res/psds-ftss/index-eng.php) | | | |
| 66. | Risk Group | RG-1  RG-2  RG-3\* | |
| 67. | Biosafety Level | BSL-1  BSL-2  BSL-2 with BSL-3 practices  BSL-3\* | |
| If you are working with **animals or plants**, use the following resources to determine the appropriate containment level for the intended use of such categories.  Note: Your work might require permitting for the transport of USDA regulated articles (genetically modified plants, soil containing plant pests, noxious weeds or pests, etc.).  BMBL: <https://www.cdc.gov/labs/pdf/CDC-BiosafetyMicrobiologicalBiomedicalLaboratories-2009-P.PDF>  NIH Guidelines: <https://osp.od.nih.gov/wp-content/uploads/2019_NIH_Guidelines.htm>  USDA/APHIS: <http://www.aphis.usda.gov/biotechnology/index.shtml>  Arthropod Containment Guidelines: <https://www.ncbi.nlm.nih.gov/pmc/articles/PMC6396570/> | | | |
| 68. | Animal Containment | | ABSL-1  ABSL-2 |
| 69. | Plant Containment | | BL1-P  BL2-P |
| 70. | Arthropod Containment | | ACL-1  ACL-2 |
| 71. | **Classification Determination**  Indicate below the relevant section(s) of the documents you used in making your determinations above: (e.g., “BMBL: Section IV - RG classification and Section VIII – Agent Summary Statements” or “NIH Guidelines: Appendix B - BSL classification”) or, if the agent(s) you are using is(are) not referenced in these documents, provide scientific rationale for your classification determinations.  Click here to enter text. | | |
| 72. | **Classification Justification**  Provide justification on why you made the specific determination(s) above:  Click here to enter text. | | |

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| **ATTESTATION** |
| By submitting this form, you (the Principal Investigator) are certifying the following: (check to indicate that you have read each one):  The information contained in this report is true, complete, and accurate to the best of your knowledge;  The research will be conducted in accordance with applicable laws, regulations, and Baylor University policies and procedures;  Research records will be kept for at least 3 years after completion of the research (a longer period may be required by a sponsor or funding agency). All records must be accessible for inspection and copying by authorized representatives.  You are aware, as the Principal Investigator, you are ultimately responsible for the conduct of this research and the individuals to whom you delegate research responsibilities. You are responsible for ensuring the training of all personnel involved in the proposed project in matters of potential biohazards, relevant biosafety practices, techniques, and emergency procedures, and that such training remains current.  You confirm that any proposed changes to the project will not be initiated or modified until appropriate approval is received. This includes the addition of personnel. |